## Total Synthesis of Calyculin C

## Anthony K. Ogawa<sup>1</sup> and Robert W. Armstrong<sup>\*,2</sup>

Contribution from the Department of Chemistry and Biochemistry, University of California, Los Angeles, California 90095

Received March 13, 1998

**Abstract:** The study of phosphatases continues to flourish given their prominence in signal transduction pathways and the regulation of cell function. Our research efforts in this area focus on the potent inhibition of serine/ threonine phosphatases, PP1 and PP2A, by a structurally diverse class of natural products. We herein report the completed total synthesis of the serine/threonine phosphatase inhibitor calyculin C (1) as part of an ongoing effort to elucidate key structural requirements for phosphatase inhibition by the aforementioned diverse class of natural products. Synthetic issues addressed include (1) the remote protecting group effect on Brown crotylboration diastereoselectivity during the introduction of the C<sub>10</sub>-C<sub>11</sub> stereocenters and (2) the formation of the C<sub>25</sub>-C<sub>26</sub> double bond using a fully deprotected C<sub>26</sub>-C<sub>37</sub> phosphonium salt (3). In addition, the concurrent synthesis of 34*R*-calyculin C (29) clarified our previous C<sub>34</sub>-stereochemical assignment of C<sub>26</sub>-C<sub>37</sub> phosphonium salts (3 and 27) (Ogawa, A. K.; DeMattei, J. A.; Scarlato, G. R.; Tellew, J. E.; Chong, L. S.; Armstrong, R. W. *J. Org. Chem.* 1996, *61*, 6153).

The study of phosphatases continues to flourish given their prominence in signal transduction pathways and the regulation of cell function.<sup>3</sup> Our research efforts in this area focus on the potent inhibition of serine/threonine phosphatases, PP1 and PP2A, by a structurally diverse class of natural products.<sup>4</sup> In a previous paper, we proposed a computationally derived enzyme inhibitor model that identified structural motifs common to all PP1 and PP2A natural product inhibitors.<sup>5</sup> (See Figure 2) Within this class of inhibitors, the calyculins<sup>6</sup> have attracted extensive synthetic interest due to their diverse functionality<sup>7–9</sup>

(1) Taken in part from the Ph.D. Thesis of Anthony K. Ogawa, University of California, Los Angeles, 1997. Present address: The Scripps Research Institute, La Jolla, CA.

(2) Present address: Amgen Inc., Thousand Oaks, CA.

(3) Wera, S.; Hemmings, B. A. *Biochem. J.* **1995**, *311*, 17, and references therein.

(4) (a) Ishihara, H.; Martin, B. L.; Brautigan, D. L.; Karaki, H.; Ozaki, H.; Kato, Y.; Fusetani, N.; Watabe, S.; Hashimoto, K.; Uemura, D. *Biochem. Biophys. Res. Commun.* **1989**, *159*, 871. (b) MacKintosh, C.; Klumpp, S. *FEBS Lett.* **1990**, *277*, 137. (c) Honkanen, R. E.; Dukelow, M.; Zwiller, J.; Moore, R. E.; Khatra, B. S.; Boynton, A. L. *Mol. Pharm.* **1991**, *40*, 577. (d) Honkanen, R. E.; Zwiller, J.; Moore, R. E.; Daily, S. L.; Khatra, B. S.; Dukelow, M.; Boynton, A. L. *J. Biol. Chem.* **1990**, *265*, 19401. (e) Dilip de Silva, E.; Williams, D. E.; Andersen, R. J.; Klix, H.; Holmes, C. F. B.; Allen, T. M. *Tetrahedron Lett.* **1992**, *33*, 1561.

(5) Gupta, V.; Ogawa, A. K.; Du, X.; Houk, K. N.; Armstrong, R. W. J. Med. Chem. **1997**, 40, 3199.

(see Figure 1). We herein report the completed total synthesis of the serine/threonine phosphatase inhibitor calyculin C (1) as part of an ongoing effort to elucidate key structural requirements for phosphatase inhibition by the aforementioned diverse class of natural products.

Retrosynthetic analysis yielded an initial disconnection at the  $C_{25}-C_{26}$  double bond that subdivided calyculin into two fragments of approximately equal functional complexity (**2** and **3**)<sup>10</sup> (see Scheme 1). Previous synthetic papers outlined a strategy for  $C_{25}-C_{26}$  double bond construction via Wittig olefination with a fully deprotected  $C_{26}-C_{37}$  phosphonium salt.<sup>7a</sup> Model studies affirmed the viability of this approach which circumvented downstream protecting group manipulations with respect to the  $C_{26}-C_{37}$  fragment synthesis. Finally, elaboration

<sup>(6) (</sup>a) Kato, Y.; Fusetani, N.; Matsunaga, S.; Hashimoto, K.; Fujita, S.; Furuya, T. J. Am. Chem. Soc. **1986**, 108, 2780. (b) Kato, Y.; Fusetani, N.; Matsunaga, S.; Hashimoto, K.; Koseki, K. J. Org. Chem. **1988**, 53, 3930. (c) Matsunaga, S.; Fujiki, H.; Sakata, D. Tetrahedron **1991**, 47, 2999. (d) Matsunaga, S.; Wakimoto, T.; Fusetani, N. J. Org. Chem. **1997**, 62, 2640. (e) Matsunaga, S.; Wakimoto, T.; Fusetani, N.; Suganuma, M. Tetrahedron Lett. **1997**, 38, 3763.

<sup>(7)</sup> Preliminary work from this laboratory: (a) Ogawa, A. K.; DeMattei, J. A.; Scarlato, G. R.; Tellew, J. E.; Chong, L. S.; Armstrong, R. W. J. Org. Chem. **1996**, 61, 6153. (b) Scarlato, G. R.; DeMattei, J. A.; Chong, L. S.; Ogawa, A. K.; Lin, M. R.; Armstrong, R. W. J. Org. Chem. **1996**, 61, 6139. (c) Zhao, Z.; Scarlato, G. R.; Armstrong, R. W. *Tetrahedron Lett.* **1992**, 33, 1609. (d) Armstrong, R. W.; DeMattei, J. A. *Tetrahedron Lett.* **1992**, 33, 5749.

<sup>(8)</sup> Previous total syntheses of calyculin A: (a) Evans, D. A.; Gage, J. R.; Leighton, J. L. *J. Am. Chem. Soc.* **1992**, *114*, 9434. (b) Tanimoto, N.; Gerritz, S. W.; Sawabe, A.; Noda, T.; Filla, S. A.; Masamune, S. Angew. Chem. **1994**, *106*, 674.

<sup>(9)</sup> For other synthetic efforts toward the calyculins: (a) Evans, D. A.; Gage, J. R.; Leighton, J. L. J. Org. Chem. 1992, 57, 1964. (b) Evans, D. A.; Gage, J. R.; Leighton, J. L.; Kim, A. S. J. Org. Chem. 1992, 57, 1961. (c) Evans, D. A.; Gage, J. R. J. Org. Chem. 1992, 57, 1958. (d) Evans, D. A.; Gage, J. R.; Tetrahedron Lett. 1990, 31, 6129. (e) Vaccaro, H. A.; Levy, D. E.; Sawabe, A.; Jaetsch, T.; Masamune, S. Tetrahedron Lett. 1992, 33, 1937. (f) Duplanter, A. J.; Nantz, M. H.; Roberts, J. C.; Short, R. P.; Somfai, P.; Masamune, S. Tetrahedron Lett. 1989, 30, 7357. (g) Kabeya, M.; Hamada, Y.; Shioiri, T. Tetrahedron 1997, 53, 9777. (h) Kabeya, M.; Hamada, Y.; Shioiri, T. Tetrahedron 1997, 53, 9769. (i) Yokokawa, F.; Hamada, Y.; Shioiri, T. Chem. Commun. 1996, 871. (j) Yokokawa, F.; Hamada, Y.; Shioiri, T. Tetrahedron Lett. 1993, 34, 6559. (k) Yokokawa, F.; Hamada, Y.; Shioiri, T. Synlett. 1992, 149. (1) Hara, O.; Hamada, Y.; Shioiri, T. Synlett. 1991, 283. (m) Smith, A. B., III; Iwashima, M. Tetrahedron Lett. 1994, 35, 6051. (n) Salvatore, B. A.; Smith, A. B., III. Tetrahderon Lett. 1994, 35, 1329. (o) Smith, A. B., III; Salvatore, B. A.; Hull, K. G.; Duan, J. J.-W. Tetrahedron Lett. 1991, 32, 4859. (p) Smith, A. B., III; Duan, J. J.-W.; Hull, K. G.; Salvatore, B. A. Tetrahedron Lett. 1991, 32, 4855. (q) Barrett, A. G. M.; Edmunds, J. J.; Hendrix, J. A.; Malecha, J. W.; Parkinson, C. J. J. Chem. Soc., Chem. Commun. 1992, 1240. (r) Barrett, A. G. M.; Edmunds, J. J.; Hendrix, J. A.; Horita, K.; Parkinson, C. J. J. Chem. Soc., Chem. Commun. 1992, 1238. (s) Barrett, A. G. M.; Edmunds, J. J.; Horita, K.; Parkinson, C. J. J. Chem. Soc., Chem. Commun. 1992, 1236. (t) Barrett, A. G. M.; Malecha, J. W. J. Org. Chem. 1991, 56, 5243. (u) Koskinen, A. M. P.; Pihko, P. M. Tetrahedron Lett. 1994, 35, 7417. (v) Koskinen, A. M. P.; Chen, J. Tetrahedron Lett. 1991, 32, 6977. (w) Trost, B. M.; Flygare, J. A. Tetrahedron Lett. 1994, 35, 4059. (10) The structure of **3** was previously assigned in ref 7a as its  $C_{34}$ epimer.

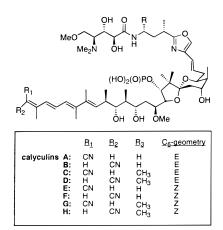


Figure 1. Scheme 1

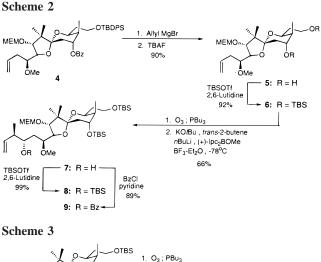
of the C<sub>17</sub>-hydroxyl late in the C<sub>1</sub>-C<sub>25</sub> fragment synthesis, as precedented by Evans and co-workers,<sup>8a</sup> imparted the requisite flexibility to explore alternate functional groups at this position. Modification at this specific location appealed due to the potential significance of the phosphate toward tertiary structure organization and/or direct active site interactions. Of interest was the apparently conflicting evidence regarding the role of the aforementioned phosphoryl moiety. Examination of the calyculin A crystal structure suggested a significant role in molecular folding, which should logically enhance activity by limiting the entropic cost upon binding. However, Fusetani recently reported that C<sub>17</sub>-desphosphoryl calyculin A was approximately as equipotent as phosphorylated calyculins,<sup>6e</sup> thus raising an interesting issue regarding the natural selection for such an unusual functionalization.

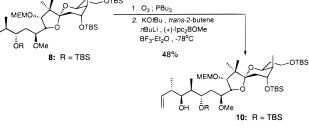
ÖR ÖR

2

оме

Prefacing the completion of the total synthesis were efforts to improve our synthesis of the  $C_1-C_{25}$  fragment (2). Targets of refinement included (1) an upstream conversion to a global silvl protecting group strategy that reduced the number of linear synthetic steps and (2) improving the selectivity during the introduction of the  $C_{10}$ , $C_{11}$ -propionate unit. The switch to a global silvl protecting group strategy proceeded from spiroketal 4 with the deprotection of both the TPS and benzoate protecting groups to afford diol 5. Silvlation yielded core structure 6 in high yield, thereby completing the protecting group exchange. Ozonolysis afforded an aldehyde intermediate that, upon





submission to an asymmetric Brown crotylboration reaction,<sup>11</sup> afforded olefin **7** as the sole reaction product. This result was consistent with the Brown algorithm that predicted an *anti*-stereochemical relationship for the C<sub>12</sub>-methyl and C<sub>13</sub>-hydroxyl given *trans*-2-butene as the carbon input. Confirmation of this assignment arose later through comparison with previously characterized intermediates<sup>7b</sup> (see Scheme 2).

The C<sub>13</sub>-hydroxyl of olefin **7** was readily protected as either a silyl ether (**8**) or benzoate ester (**9**). Ozonolysis of olefin **8**, again followed by Brown crotylboration,<sup>11</sup> yielded a single alkylated reaction product (**10**). The drastic change in diastereoselectivity from previous results on this section of calyculin raised suspicion regarding the identity of the newly generated  $C_{10}$ , $C_{11}$  stereocenters (see Scheme 3).

To assign the newly generated stereocenters, acetonide formation on the C11,C13-diol was effected to utilize the Rychnovsky correlation for deducing 1,3-diol stereochemical relationships.<sup>12</sup> The determination of stereochemistry followed from the comparison of the <sup>13</sup>C chemical shifts for the 1,3-Oacetonide methyl groups of the 1,3-dioxane system, which has been shown to be generally independent of C<sub>2</sub>-substitution. To this end, olefin 10 was fully desilylated to afford tetrol 11. The reaction of tetrol 11 in 2,2-dimethoxypropane in the presence of catalytic acid afforded acetonides 12 and 13, the latter of which could be readily converted to the desired mono-ketal product via mild acid hydrolysis. Analysis of the <sup>13</sup>C spectrum for 12 confirmed an anti-stereochemical relationship based on the observed resonances, 23.7 and 24.5 ppm. Evidently, the  $\beta$ -hydroxyl protecting group played a significant role in the asymmetric induction at C11 that completely overrode the induction predicted for the Brown crotylboration!<sup>11</sup> Indeed, the overwhelming preference for the anti-syn-anti relationship

<sup>(11)</sup> Representative references: (a) Brown, H. C.; Bhat, K. S. J. Am. Chem. Soc. **1986**, 108, 293. (b) Brown, H. C.; Jadhav, P. K.; Bhat, K. S. J. Am. Chem. Soc. **1988**, 110, 1535.

<sup>(12)</sup> Rychnovsky, S. D.; Rogers, B.; Yang, G. J. Org. Chem. 1993, 58, 3511.

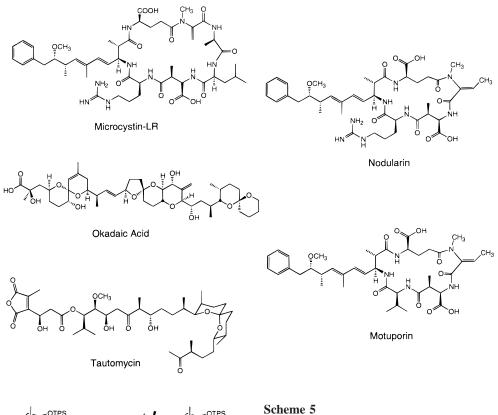


Figure 2.

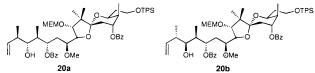
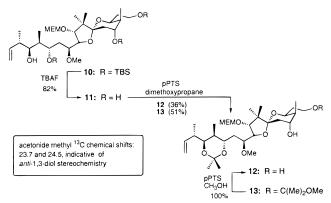


Figure 3.

Scheme 4



observed for  $C_{10}$ - $C_{13}$  in **10** was consistent with the closed transition state model of Roush<sup>13</sup> (see Scheme 4).

This result represented a drastic departure from our previously reported synthesis of the  $C_1-C_{25}$  fragment, in which the *anti-anti-anti* isomer was preferentially generated by a 40:30 diastereomeric ratio (**20a:20b**, respectively)<sup>7b</sup> (see Figure 3). The aldehyde used in that Brown crotylboration was benzoyl-protected at  $C_{13}$ , which prompted the submission of olefin **9** to a similar ozonolysis-Brown crotylboration sequence. This yielded the desired *anti-anti-anti* diastereomeric mixture (**14a** and **14b**) favoring the *anti-syn-anti* isomer. Positive structural assignments were made through the conversion of each dia-

stereomer to their corresponding tetra-TBS derivatives (**21** and **22**) and comparison to previously reported compound<sup>7b</sup> (see Scheme 7).

OTBS

ÓTBS

ōвz

1. O3; PBu3

KO/Bu , trans-2-butene

ÕH ÖBz

ME

ĎВz

ŌΜε

OTBS

OTBS

ÓTBS

14a (22%)

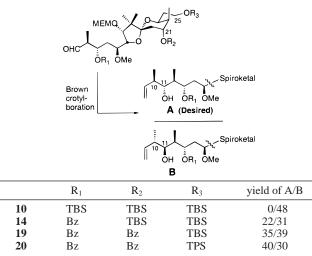
14b (31%)

nBuLi , (+)-lpc<sub>2</sub>BOMe BF<sub>3</sub>-Et<sub>2</sub>O , -78°C

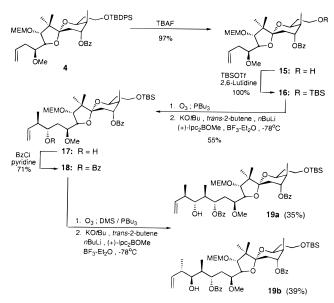
Despite the obvious effect of the  $\beta$ -alkoxy protecting group on stereoselectivity, the discrepancy between the diastereomeric ratios of 14a:14b and 20a:20b suggested that the remote protecting groups at C<sub>21</sub> and C<sub>25</sub> also impacted the crotylboration diastereoselectivity. This subtle steric effect arose from the *R*-stereochemistry at  $C_{16}$ , which projected the reaction site ( $C_{11}$ ) toward the aforementioned alkoxy groups (protected C<sub>21</sub> and  $C_{25}$ ). Illustration of this point derived from the effect of a  $C_{25}$ hydroxyl TBS for TPS exchange on diastereoselectivity at  $C_{10}$ ,  $C_{11}$ . Spiroketal 4 was converted to its  $C_{25}$ -TBS protected derivative (16) in excellent yield. An initial ozonolysis-Brown crotylboration sequence yielded, as expected, a single reaction product (17), the C<sub>13</sub>-hydroxyl of which was benzoyl-protected (18). The subsequent ozonolysis-Brown crotylboration sequence led to a modest reversal of distereoselectivity to yield homoallylic alcohols 19a and 19b in a 35:39 ratio, respectively. Clearly, a definite and remarkable trend exists in which variation

<sup>(13)</sup> Roush, W. R. J. Org. Chem. 1991, 56, 4151.

Table 1



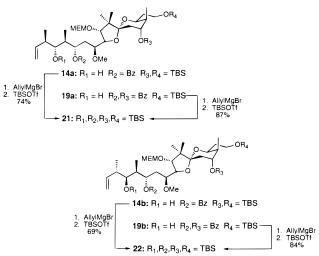
Scheme 6



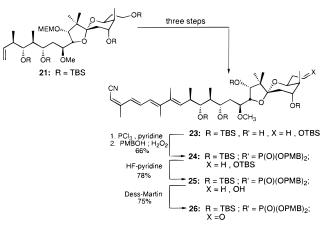
of proximal and remote protecting groups affected the crotylboration diastereoselectivity! (see Table 1)

The conversion of *anti-anti-anti* olefin isomers (**14a** and **19a**) to a common tetra-TBS protected compound (**21**) enabled the conclusive assignment of stereochemistry at  $C_{10}-C_{13}$  and established the desired global silyl protection scheme. Elaboration to tetraene **23** over four steps followed a previously established route.<sup>7b,8a</sup> Phosphorylation afforded **24** in good yield using the Evans procedure<sup>8a</sup> (see Scheme 2). Selective deprotection of the C<sub>25</sub>-TBS group (**25**), followed by Dess–Martin oxidation of the corresponding primary hydroxyl<sup>14</sup> provided aldehyde **26** and completed the C<sub>1</sub>–C<sub>25</sub> fragment synthesis (see Scheme 8).

In our previously reported synthesis of the  $C_{26}-C_{37}$  fragment,<sup>7a</sup> Weinreb amidation<sup>15</sup> to generate the  $C_{33}-N_3$  amide bond resulted in a  $C_{34}$ -epimeric mixture of products,<sup>16</sup> which we were unable to definitively assign. Consequently, both diastereomers were carried on into coupling reactions with the  $C_1-C_{25}$  Scheme 7



Scheme 8



fragment. Condensation of the deprotected phosphonium salt derived from the major amide stereoisomer with aldehyde **26** proceeded in good yield. Submission of the corresponding Wittig adduct to HF-deprotection afforded a TLC spot that failed to coelute with an authentic sample of calyculin C. <sup>1</sup>H NMR and HRMS analysis of the resultant product confirmed its identity as a fully deprotected system. This evidence supported our structural assignment of **29** as  $C_{34}$ -*epi*-calyculin C and the major Weinreb amidation product as stereoisomer **27** (see Scheme 9).

Condensation of the phosphonium salt from the minor amidation product (**3**) with aldehyde **26** proceeded in acceptable yield to protected calyculin **30**. Deprotection of **30** under the previously employed reaction conditions generated a TLC spot that co-spotted with an authentic sample in multiple solvent systems.<sup>17</sup> <sup>1</sup>H NMR<sup>18</sup> and HRMS data were consistent with the aforementioned authentic calyculin C sample, which corroborated our structural assignments of compounds **27–30** and represented the completed total synthesis of calyculin C (**1**) (see Scheme 10).

In summary, our pursuit of an improved synthesis for the  $C_1-C_{25}$  fragment exposed a remarkable, remote protecting group effect on the diastereoselectivity that was observed during the

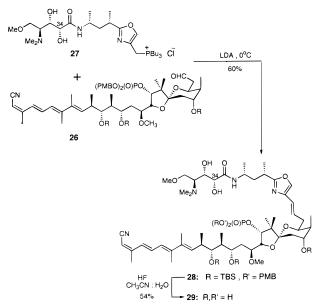
<sup>(14)</sup> Meyer, S. D.; Schreiber, S. L. J. Org. Chem. 1994, 59, 7549.

<sup>(15)</sup> Basha, A.; Lipton, M.; Weinreb, S. M. *Tetrahedron Lett.* **1977**, 4171. (16) Noteworthy is our ability to recycle recovered ester from the Weinreb amidation. The diastereoselectivity of the amidation is not affected by the corresponding epimeric mixture present in the ester starting material.

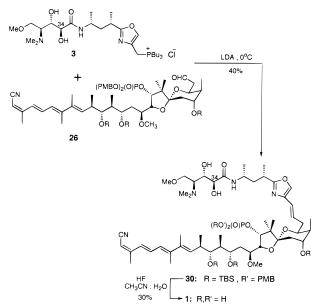
<sup>(17)</sup> The solvent systems sampled were (a) 10% acetone:ethyl acetate ( $R_f = 0.7$ ), (b) 30% acetone:hexane (double elution;  $R_f = 0.3$ ), (c) 20% acetonitrile:methylene chloride (double elution;  $R_f = 0.6$ ), and (d) 5% methanol:methylene chloride (double elution;  $R_f = 0.4$ ).

<sup>(18) 1</sup>H NMR data for authentic and synthetic calyculin C were obtained in both CDCl<sub>3</sub> and  $C_6D_6$ .

Scheme 9



Scheme 10



generation of the  $C_{10}$ ,  $C_{11}$ -propionate stereocenters. In addition, we were able to unequivocally assign the stereochemistry at  $C_{34}$ . We, therefore, report the total synthesis of calyculin C, a member of a family of structurally diverse serine/threonine phosphatase inhibitors.

## **Experimental Section**

**General Methods.** NMR spectra were obtained from Bruker AM360, ARX400, and ARX500 spectrometers. IR data was collected on a Nicolet PCIR. Optical rotations were taken at 22 °C on a Perkin-Elmer model 241MC. High-resolution FAB mass spectra were obtained by the Mass Spectroscopy Facilities at UCLA or UC-Riverside. For FAB mass spectra,  $2\sigma = 4$  ppm.

Solvents and reagents were used as supplied from commercial sources with the following exceptions. All moisture sensitive reagents not packaged in Aldrich sure seal bottles were distilled prior to use and stored in a desiccated environment. Particularly sensitive reagents, such as dimethyl phosphite and phosphorus trichloride, were freshly distilled prior to each usage. Tetrahydrofuran and diethyl ether were distilled from sodium benzophenone ketyl immediately prior to use. Dichloromethane was distilled from phosphorus pentoxide. Methanol was distilled from magnesium turnings. Dimethylformamide, dimethyl sulfoxide, and in some cases diisopropylamine were distilled from barium oxide and stored over 4 Å molecular sieves. All reactions involving moisture-sensitive reagents were performed under either a nitrogen or an argon atmosphere.

(2*R*,3*R*,5*R*,7*S*,8*R*,9*R*)-9-Hydroxy-7-(2-hydroxyethyl)-2-[(1*S*)-methoxy-but-3ene]-3-(2-methoxyethoxymethyl)oxy-4,4,8-trimethyl-1,6dioxaspiro[4.5]decane (5). To a solution of benzoate 4 (1.60 g, 2.06 mmol, 1.0 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (45 mL) at -78 °C was added allylmagnesium bromide (1.0 M in diethyl ether, 15 mL, 7.25 equiv) rapidly dropwise. The cloudy white reaction mixture was warmed to room temperature over 30 min. The reaction was quenched via addition of H<sub>2</sub>O (10 mL), and the resulting suspension was filtered. The solid residue was washed with CH<sub>2</sub>Cl<sub>2</sub> (3 × 40 mL), and the filtrate was partitioned between CH<sub>2</sub>Cl<sub>2</sub> (20 mL) and brine (60 mL). The layers were separated, and the aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 40 mL). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated.

The resulting crude, yellow oil was eluted in THF (50 mL), and TBAF (1.0 M in THF, 2.5 mL, 1.2 equiv) was added. After 16 h, the reaction mixture was concentrated, and column chromatography on silica gel (70–100% ethyl acetate—hexane) afforded diol **5** (804 mg, 90%). [ $\alpha$ ]<sub>D</sub> = -64.5 (*c* 5.0, CHCl<sub>3</sub>). IR (thin film) 3501, 2924, 1472, 1419 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  5.87 (1H, m), 5.10 (1H, dd, *J* = 17.2, 1.2 Hz), 5.06 (1H, m), 4.71 (2H, s), 4.47 (1H, dt, *J* = 11.2, 2.0 Hz), 4.16 (1H, dd, *J* = 9.1, 4.9 Hz), 4.06 (1H, dd, *J* = 11.1, 1.1 Hz), 3.80 (2H, m), 3.71 (4H, m), 3.52, (3H, s), 3.48 (2H, m), 3.33 (3H, s), 2.47 (1H, m), 1.57 (1H, m, J = 14.2 Hz), 1.38 (1H, m), 1.06 (3H, s), 0.90 (3H, s), 0.84 (3H, d, *J* = 7.1 Hz). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  134.1, 117.3, 108.2, 97.3, 86.7, 83.4, 80.0, 71.5, 70.5, 69.9, 68.5, 63.5, 59.0, 58.9, 50.5, 38.1, 34.8, 34.4, 28.8, 22.9, 17.5, 10.8. HRFABMS calcd for (M + H) C<sub>22</sub>H<sub>41</sub>O<sub>8</sub>: 433.2801, found: 433.2807.

(2R,3R,5R,7S,8R,9R)-9-tert-Butyldimethylsiloxy-7-(2-tert-butyldimethylsiloxyethyl)-2-[(1S)-methoxy-but-3ene]-3-(2-methoxyethoxymethyl)oxy-4,4,8-trimethyl-1,6-dioxaspiro[4.5]decane (6). To a solution of diol 5 (50 mg, 0.116 mmol, 1.0 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) at 0 °C was added 2,6-lutidene (81 µL, 0.693 mmol, 6 equiv). TBSOTf (80 µL, 0.347 mmol, 3 equiv) was added dropwise, and the clear reaction mixture was stirred at 0 °C for 15 min. Saturated aqueous NaHCO3 (2 mL) was added, and the resulting mixture was partitioned between CH<sub>2</sub>Cl<sub>2</sub> (20 mL) and brine (15 mL). The layers were separated, and the aqueous layer was extracted with  $CH_2Cl_2$  (2 × 30 mL). The combined organics were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. Purification via column chromatography on silica gel (10% ethyl acetate-hexane) afforded silvl ether 6 (70 mg, 92%).  $[\alpha]_D = -75.8$ (c 3.6, CHCl<sub>3</sub>). IR (thin film) 2928, 1471, cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz,  $CDCl_3$ )  $\delta$  5.87 (1H, m), 5.07 (1H, dd, J = 17.2, 1.7 Hz), 5.00 (1H, m), 4.70 (1H, d, J = 13.1 Hz), 4.67 (1H, d, J = 13.1 Hz), 4.34 (1H, dt, J = 10.6, 2.3 Hz), 4.02 (1H, dd, J = 9.2, 5.2 Hz), 3.90 (1H, ddd, J =15.5, 10.4, 5.1 Hz), 3.78 (1H, m), 3.62-3.70 (4H, m), 3.59 (3H, s), 3.49 (2H, m), 3.38 (1H, dd, J = 9.3, 2.3 Hz), 3.35 (3H, s), 2.34 (1H, dd, J = 9.3, 2.3 Hz), 3.35 (3H, s), 2.34 (1H, dd, J = 9.3, 2.3 Hz), 3.35 (3H, s), 2.34 (1H, dd, J = 9.3, 2.3 Hz), 3.35 (3H, s), 2.34 (1H, dd, J = 9.3, 2.3 Hz), 3.35 (3H, s), 2.34 (1H, dd, J = 9.3, 2.3 Hz), 3.35 (3H, s), 2.34 (1H, dd, J = 9.3, 2.3 Hz), 3.35 (3H, s), 2.34 (1H, dd, J = 9.3, 2.3 Hz), 3.35 (2H, s), 3.35 (2H, s)m), 2.04 (1H, m), 1.72 (1H, m), 1.60 (1H, dd, *J* = 14.3, 3.8 Hz), 1.36– 1.44 (3H, m), 1.00 (3H, s), 0.88 (9H, s), 0.85 (9H, s), 0.78 (3H, d, J = 7.1 Hz), 0.04 (3H, s), 0.03 (3H, s), 0.02 (3H, s), -0.02 (3H, s).  $^{13}C$ NMR (125 MHz, CDCl<sub>3</sub>) δ 135.6, 116.1, 106.8, 97.6, 87.7, 84.1, 81.0, 71.6, 71.2, 68.1, 63.5, 62.1, 60.4, 59.0, 50.5, 38.7, 36.5, 35.7, 30.6, 26.1, 25.8, 23.0, 18.4, 18.1, 17.7, 10.5, -4.7, -5.1, -5.2 (2 carbons). HRFABMS calcd for (M + H) C<sub>34</sub>H<sub>69</sub>O<sub>8</sub>Si<sub>2</sub>: 661.4531, found: 661.4529

(2R,2(1S,3S,4S),3R,5R,7S,8R,9R)-9-tert-Butyldimethylsiloxy-7-(2tert-butyldimethylsiloxyethyl)-2-(3-hydroxy-1-methoxy-4-methyl-5hexenyl)-3-(2-methoxyethoxymethyl)oxy-4,4,8-trimethyl-1,6-dioxaspiro[4.5]decane (7). Ozone was bubbled through a solution of olefin 6 (993 mg, 1.51 mmol, 1.0 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (15 mL) at -78 °C until a light blue endpoint was achieved. The excess ozone was purged via a stream of argon (until colorless). Tributylphosphine (350  $\mu$ L, 1.47 mmol, 0.97 equiv) was added dropwise, and the clear colorless reaction mixture was warmed slowly to room temperature. After 12.5 h, the reaction mixture was concentrated to afford a crude aldehyde which was carried on without purification. It should be noted that, although the aldehyde generally withstood column chromatography, purification did not improve the yields of the subsequent crotylboration reactions.

To a cloudy solution of KOtBu (676 mg, 6.02 mmol, 4.0 equiv) and trans-2-butene (~1 mL, excess) in THF (9 mL) at -78 °C was added dropwise 2.14 M nBuLi (2.4 mL, 5.13 mmol, 3.4 equiv). Upon initial addition of nBuLi, the reaction mixture became yellow. After, 15 min, a solution of (+)-B-Methoxydiisopinocampheylborane (2.18 g, 6.89 mmol, 4.6 equiv) in THF (2.5 mL) was added dropwise. Addition of the borane solution resulted in a clear colorless reaction mixture. After 30 min, BF<sub>3</sub>-Et<sub>2</sub>O (820 µL, 6.66 mmol, 4.4 equiv) was added rapidly, followed immediately by a solution of the crude aldehyde in THF (2 mL + 0.7 mL rinse). The resulting cloudy, colorless reaction mixture was stirred at -78 °C for 4 h. Then 3 N NaOH (3 mL) and 30% H<sub>2</sub>O<sub>2</sub> (1.5 mL) were added, and the cold bath was removed. After 3 h, the mixture was diluted with ethyl acetate (20 mL), poured into brine (20 mL), and the layers were separated. The aqueous layer was extracted with ethyl acetate (3  $\times$  40 mL), and the combined organics were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated. Purification via column chromatography on silica gel (5% ethyl acetate-hexane) afforded olefin 7 (714 mg, 66%).  $[\alpha]_D = -61.7$  (c 7.7, CHCl<sub>3</sub>). IR (thin film) 3503, 2928, 1472, cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$ 5.83 (1H, ddd, J = 18.6, 10.7, 7.8 Hz), 5.05 (1H, s), 5.03 (1H, m), 4.70 (1H, d, J = 6.4 Hz), 4.63 (1H, d, J = 6.4 Hz), 4.36 (1H, m), 4.12 (1H, dd, J = 9.4, 5.1 Hz), 3.88 (1H, m), 3.77–3.80 (2H, m), 3.62– 3.72 (6H, m), 3.51 (3H, m), 3.36 (3H, s), 2.82 (1H, d, J = 5.0 Hz), 2.18 (1H, m), 1.70 (1H, m), 1.57-1.64 (2H, m), 1.38-1.48 (4H, m), 1.02 (3H, d, J = 7.0 Hz), 1.01 (3H, s), 0.89 (9H, s), 0.88 (9H, s), 0.80(3H, d, J = 7.2 Hz), 0.04 (3H, s), 0.03 (3H, s), 0.03 (3H, s), -0.01(3H, s). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  141.0. 115.0, 106.9, 98.1, 88.3, 84.3, 79.4, 71.8, 71.6, 71.2, 68.0, 63.5, 61.8, 60.6, 59.0, 50.5, 44.4, 38.6, 36.5, 35.4, 30.6, 26.0, 25.8, 23.0, 18.3, 18.2, 17.6, 16.0, 10.5, -4.7, -5.1, -5.2, -5.2. HRFABMS calcd for (M + H) C37H75O9Si2: 719.4949, found: 719.4953.

(2R,2(15,35,45),3R,5R,75,8R,9R)-9-tert-Butyldimethylsiloxy-7-(2tert-butyldimethylsiloxyethyl)-2-(1-methoxy-4-methyl-3-(benzoyl)oxy-5-hexenyl)-3-(2-methoxyethoxymethyl)oxy-4,4,8-trimethyl-1,6-dioxaspiro[4.5]decane (9). To a solution of alcohol 7 (99 mg, 0.138 mmol, 1 equiv) in pyridine (2 mL) was added benzoyl chloride (32  $\mu$ L, 0.276 mmol, 2.0 equiv). The clear yellow mixture was heated to 50 °C for 24 h. Concentration in vacuo, followed by purification via column chromatography on silica gel (0-5% ethyl acetate/(1:1) methylene chloride/hexane) afforded spirocycle 9 (101 mg, 89%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.02 (2H, d, J = 7.4 Hz), 7.52 (1H, m), 7.41 (2H, m), 5.84 (1H, ddd, J = 17.6, 9.7, 7.4 Hz), 5.48 (1H, m), 5.03 (2H, m), 4.70 (1H, d, J = 6.9 Hz), 4.65 (1H, d, J = 6.9 Hz), 4.31 (1H, m), 4.01 (1H, dd, J = 8.9, 4.9 Hz), 3.78 (1H, d, J = 2.3 Hz), 3.70 (1H, m), 3.55-3.65 (6H, m), 3.43-3.50 (3H, m), 3.36 (4H, m), 2.52 (1H, m), 1.78 (1H, dd, J = 13.1, 10.8 Hz), 1.60 (1H, dd, J = 14.2, 3.7 Hz), 1.45-1.56 (2H, m), 1.30-1.42 (3H, m), 1.05 (3H, d, J = 8.6Hz), 1.02 (3H, s), 0.84 (12H, m), 0.76 (12H, m), 0.02 (3H, s), -0.03 (3H, s), -0.15 (3H, s), -0.16 (3H, s).  $^{13}\text{CNMR}$  (125 MHz, CDCl3)  $\delta$ 166.1, 139.3, 132.6, 130.8, 129.6, 128.2, 115.6, 106.8, 98.0, 88.1, 84.9, 77.9, 74.0, 71.7, 71.1, 67.9, 63.8, 61.1, 60.9, 59.0, 50.4, 42.5, 38.1, 34.1, 30.7, 25.8, 25.8, 23.2, 18.1, 18.0, 17.8, 15.8, 10.4, -4.7, -5.2, -5.4, -5.4, -5.4.

(2R,2(1S,3S,4S,5R,6R),3R,5R,7S,8R,9R)-9-tert-Butyldimethylsiloxy-7-(2-tert-butyldimethylsiloxyethyl)-2-(5-hydroxy-1-methoxy-4,6-dimethyl-3-benzoyloxy-8-nonenyl)-3-(2-methoxyethoxymethyl)oxy-4,4,8-trimethyl-1,6-dioxaspiro[4.5]decane (14a). Ozone was bubbled through a solution of olefin 9 (500 mg, 0.609 mmol, 1 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (25 mL) at -78 °C until a light blue color persisted. The excess ozone was purged via a stream of argon (until colorless), at which time DMS (2.3 mL, >50 equiv) was added. The reaction mixture was warmed to room temperature slowly. After 14 h, triphenylphosphine (84 mg total, ~0.6 equiv) was added in three portions over 3 h. The reaction mixture was concentrated after 19 h, and the resulting crude aldehyde was carried on without purification.

To a cloudy solution of KOtBu (352 mg, 3.14 mmol, 5.1 equiv) and *trans*-2-butene ( $\sim$ 1 mL, excess) in THF (4 mL) at -78 °C was added dropwise 2.02 M *n*BuLi (1.35 mL, 2.74 mmol, 4.5 equiv). Upon

initial addition of *n*BuLi, the reaction mixture became yellow. After, 15 min, a solution of (+)-B-Methoxydiisopinocampheylborane (1.03 g, 3.26 mmol, 5.4 equiv) in THF (1.2 mL) was added dropwise. Addition of the borane solution resulted in a clear colorless reaction mixture. After 30 min, BF<sub>3</sub>-Et<sub>2</sub>O (350 µL, 2.85 mmol, 4.7 equiv) was added rapidly, followed immediately by a solution of the crude aldehyde from above in THF (1.2 mL + 0.5 mL rinse). The resulting cloudy, colorless reaction mixture was stirred at -78 °C for 4 h. Then 3 N NaOH (4 mL) and 30% H<sub>2</sub>O<sub>2</sub> (2 mL) were added, and the cold bath was removed. After 13 h, the mixture was diluted with ethyl acetate (20 mL) and poured into brine (20 mL), and the layers were separated. The aqueous layer was extracted with ethyl acetate (3  $\times$  25 mL), and the combined organics were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The resultant crude was eluted in THF (8 mL), 3 N NaOH (2 mL), and 30% H<sub>2</sub>O<sub>2</sub> (1 mL) to complete oxidation of the borane intermediate. After 2 h, the mixture was worked-up as previously mentioned. The excess isopinocamphol was removed via Kugelrohr distillation (high vacuum pressure at 80 °C). Purification via column chromatography on silica gel (5-20% ethyl acetate-hexane) afforded olefins 14a (118 mg, 22%) and 14b (163 mg, 31%) (53% overall for both steps). For 14a:  $[\alpha]_D = -59.1$  (c 2.4, CHCl<sub>3</sub>). IR (thin film) 3443, 2928, 1719, 1377 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.04 (2H, m), 7.54 (1H, m), 7.42 (2H, m), 5.87 (1H, ddd, J = 17.1, 8.6, 6.6 Hz), 5.64 (1H, m), 5.06 (2H, m), 4.70 (1H, d, J = 6.8 Hz), 4.65 (1H, d, J = 6.8 Hz), 4.34 (1H, m), 4.05 (1H, dd, J = 9.0, 4.9 Hz), 3.80 (1H, m), 3.69 (3H, s), 3.35–3.75 (9H, m), 3.33 (3H, s), 2.96 (1H, br d, J = 5.1 Hz), 4.41 (1H, m), 2.12 (1H, ddd, J = 9.0, 6.9, 3.4 Hz), 1.77 (2H, m), 1.62 (1H, m), 1.42 (3H, m), 1.16 (3H, d, J = 6.9 Hz), 1.04 (3H, s), 0.95 (3H, d, J = 6.9 Hz), 0.89 (3H, s), 0.88 (3H, s), 0.86 (9H, s), 0.80 (9H, s), 0.79 (3H, d, J = 5.1 Hz), 0.03 (3H, s), -0.01 (3H, s),-0.09 (6H, s). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 165.8, 139.3, 132.7, 130.9, 129.6, 128.3, 115.8, 107.0, 98.1, 88.1, 84.9, 79.2, 76.6, 73.3, 71.7, 71.2, 68.0, 63.8, 61.3, 60.9, 59.0, 50.4, 40.6, 39.6, 38.3, 36.3, 32.2, 30.8, 29.7, 26.1, 25.9 (two carbons), 23.2, 18.2, 18.2, 18.0, 17.8, 11.6, 10.5, -4.6, -5.1, -5.3 (two carbons). HRFABMS calcd for (M + Na) C<sub>47</sub>H<sub>84</sub>O<sub>11</sub>Si<sub>2</sub>Na: 903.5450, found: 903.5459.

(2R,2(15,35,45,55,65),3R,5R,75,8R,9R)-9-tert-Butyldimethylsiloxy-7-(2-tert-butyldimethylsiloxyethyl)-2-(5-hydroxy-1-methoxy-4,6-dimethyl-3-benzoyloxy-8-nonenyl)-3-(2-methoxyethoxymethyl)oxy-**4,4,8-trimethyl-1,6-dioxaspiro**[**4.5**]**decane** (14b).  $[\alpha]_D = -57.7$  (*c* 3.4, CHCl<sub>3</sub>). IR (thin film) 3492, 2928, 1717, 1472 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.05 (2H, m), 7.56 (1H, m), 7.43 (2H, m), 5.85 (1H, ddd, J = 17.6, 10.3, 7.9 Hz), 5.47 (1H, ddd, J = 9.5, 6.9, 2.2 Hz), 5.08 (2H, m), 4.76 (1H, d, *J* = 6.9 Hz), 4.72 (1H, d, *J* = 6.9 Hz), 4.31 (1H, ddd, J = 7.9, 5.2, 2.0 Hz), 4.06 (1H, dd, J = 9.0, 5.0 Hz), 3.79 (1H, d, J = 2.8 Hz), 3.74 (2H, m), 3.55 (3H, s), 3.37 (3H, s),3.35-3.65 (7H, m), 2.64 (1H, d, J = 3.7 Hz), 2.32 (1H, m), 1.95 (2H, m), 1.76 (1H, m), 1.63 (1H, dd, J = 14.3, 3.9 Hz), 1.51 (1H, m), 1.41 (3H, m), 1.04 (3H, s), 0.98 (3H, d, J = 6.9 Hz), 0.94 (3H, d, J = 6.8 Hz), 0.87 (3H, s), 0.82 (9H, s), 0.79 (9H, s), 0.78 (3H, d, *J* = 7.2 Hz), 0.02 (3H, s), -0.02 (3H, s), -0.11 (3H, s), -0.12 (3H, s). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 166.9, 141.9, 132.9, 130.4, 129.7, 128.4, 115.1, 106.9, 98.0, 87.7, 84.9, 78.2, 74.4, 73.8, 71.8, 71.2, 68.1, 64.0, 61.1, 60.9, 59.0, 50.5, 41.0, 39.4, 38.0, 36.0, 33.9, 30.7, 26.1, 25.9, 25.9, 23.2, 18.2, 18.1, 17.8, 16.5, 10.4, 8.7, -4.7, -5.1, -5.3, -5.3. HRFABMS calcd for  $(M + H) C_{47}H_{85}O_{11}Si_2$ : 881.5630, found: 881.5630.

(2*R*,3*R*,5*R*,7*S*,8*R*,9*R*)-9-Benzoyloxy-7-(2-hydroxyethyl)-2-[(1*S*)methoxy-but-3ene]-3-(2-methoxyethoxymethyl)oxy-4,4,8-trimethyl-1,6-dioxaspiro[4.5]decane (15). To a solution of silyl ether 4 (1.100 g, 1.42 mmol, 1 equiv) in THF (15 mL) was added 1.0 M TBAF (3.5 mL, 3.55 mmol, 2.5 equiv). After 2.5 h, the reaction mixture was concentrated. Purification via column chromatography on silica gel (30–50% ethyl acetate—hexane) afforded alcohol **15** (736 mg, 97%). [ $\alpha$ ]<sub>D</sub> = -52.0 (*c* 28.0, CHCl<sub>3</sub>). IR (thin film) 3497, 2924, 1713, 1453 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.13 (2H, m), 7.49 (1H, m), 7.37 (2H, m), 5.87 (1H, m), 5.11 (1H, m), 5.07 (1H, dd, *J* = 17.2, 1.0 Hz), 5.02 (1H, d, 10.0 Hz), 4.71 (2H, s), 4.58 (1H, m), 4.19 (1H, m), 4.14 (1H, dd, *J* = 9.1, 4.8 Hz), 3.83 (1H, m), 3.65–3.74 (4H, m), 3.53 (4H, m), 3.46 (2H, m), 3.30 (3H, s), 2.43 (1H, m), 2.15 (1H, m), 1.89 (1H, m), 1.84 (1H, dd, *J* = 15.0, 4.1 Hz), 1.74 (2H, m), 1.35 (1H, dd, J = 14.5, 4.0 Hz, 1.08 (3H, s), 0.97 (3H, d, J = 7.1 Hz), 0.88 (3H, s).<sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  165.9, 134.6, 132.7, 130.6, 129.8, 128.1, 116.7, 106.6, 97.2, 87.2, 83.1, 80.3, 73.4, 71,4, 69.9, 68.3, 63.0, 59.1, 58.8, 50.6, 35.2, 34.8, 34.7, 27.3, 22.7, 17.4, 10.2. HRFABMS calcd for (M + H) C<sub>29</sub>H<sub>45</sub>O<sub>9</sub>: 537.3064, found: 537.3066.

(2R,3R,5R,7S,8R,9R)-9-Benzoyloxy-7-(2-tert-butyldimethylsiloxyethyl)-2-[(1S)-methoxy-but-3ene]-3-(2-methoxyethoxy methyl)oxy-4,4,8-trimethyl-1,6-dioxaspiro[4.5]decane (16). To a solution of alcohol 15 (736 mg, 1.37 mmol, 1 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (15 mL) at 0 °C was added 2,6-lutidine (390 µL, 3.34 mmol, 2.4 equiv), followed by TBSOTf (380  $\mu$ L, 1.65 mmol, 1.2 equiv). After 10 min, the reaction mixture was partitioned between CH2Cl2 (30 mL) and saturated aqueous NaHCO<sub>3</sub> (20 mL). The layers were separated, and the aqueous layer was extracted with  $CH_2Cl_2$  (3 × 30 mL). The combined organics were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. Purification via column chromatography on silica gel (5-15% ethyl acetate-hexane) afforded silyl ether 16 (924 mg, 103%, contaminated with TBS-OH).  $[\alpha]_D =$ -77.7 (c 20.2, CHCl<sub>3</sub>). IR (thin film) 2928, 1715, 1453 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.14 (2H, m), 7.48 (1H, m), 7.38 (2H, m), 5.88 (1H, dddd, J = 17.7, 7.8, 4.3, 2.1 Hz), 5.13 (1H, m), 5.07 (1H, dd, J = 17.1, 1.2 Hz), 5.01 (1H, d, J = 10.2 Hz), 4.69 (2H, m), 4.42 (1H, m), 4.07 (1H, dd, J = 9.3, 5.2 Hz), 3.90 (1H, m), 3.68 (3H, m), 3.62 (1H, d, J = 5.2 Hz), 3.51 (3H, s), 3.49 (5H, m), 3.33 (3H, s),2.40 (1H, m), 2.11 (1H, m), 1.81 (1H, dd, J = 11.1, 4.0 Hz), 1.68– 1.78 (3H, m), 1.46 (1H, m), 1.07 (3H, s), 0.94 (3H, d, J = 7.1 Hz), 0.86 (3H, s), 0.84 (9H, s), 0.01 (3H, s), 0.00 (3H, s). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 165.9, 135.1, 132.5, 130.7, 129.8, 128.0, 116.2, 106.3, 97.5, 87.2, 83.1, 80.6, 72.9, 71.5, 68.0, 63.8, 61.4, 59.2, 58.8, 50.5, 36.0, 34.9, 34.8, 27.5, 25.9, 22.8, 18.2, 17.5, 10.0, -5.4, -5.4. HRFABMS calcd for (M + Na)  $C_{35}H_{58}O_9SiNa$ : 673.3748, found: 673.3775.

(2*R*,2(1*S*,3*S*,4*S*),3*R*,5*R*,7*S*,8*R*,9*R*)-9-Benzoyloxy-7-(2-*tert*-butyldimethylsiloxyethyl)-2-(3-hydroxy-1-methoxy-4-methyl-5-hexenyl)-3-(2-methoxyethoxymethyl)oxy-4,4,8-trimethyl-1,6-dioxaspiro[4.5]decane (17). Ozone was bubbled through a solution of olefin 16 (653 mg, 1.00 mmol, 1.0 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (12 mL) at -78 °C until a light blue endpoint was achieved. The excess ozone was purged via a stream of argon (until colorless). Tributylphosphine (250  $\mu$ L, 1.05 mmol, 1.05 equiv) was added dropwise, and the clear, colorless reaction mixture was warmed slowly to room temperature. After 10 h, the reaction mixture was concentrated to afford a crude aldehyde that was carried on without purification.

To a cloudy solution of KOtBu (560 mg, 4.99 mmol, 5.0 equiv) and trans-2-butene (~1.5 mL, excess) in THF (6 mL) at -78 °C was added dropwise 2.02 M nBuLi (2.3 mL, 4.65 mmol, 4.6 equiv). Upon initial addition of *n*BuLi, the reaction mixture became yellow. After, 15 min, a solution of (+)-B-Methoxydiisopinocampheylborane (1.72 g, 5.42 mmol, 5.4 equiv) in THF (2 mL) was added dropwise. Addition of the borane solution resulted in a clear, colorless reaction mixture. After 30 min, BF<sub>3</sub>-Et<sub>2</sub>O (590 µL, 4.79 mmol, 4.8 equiv) was added rapidly, followed immediately by a solution of the crude aldehyde in THF (2.4 mL + 1 mL rinse). The resulting cloudy colorless reaction mixture was stirred at -78 °C for 3 h. Then 3 N NaOH (4 mL) and 30% H<sub>2</sub>O<sub>2</sub> (2 mL) were added, and the cold bath was removed. After 5 h, the mixture was diluted with ethyl acetate (50 mL) and poured into brine (20 mL), and the layers were separated. The aqueous layer was extracted with ethyl acetate (4  $\times$  50 mL), and the combined organics were dried over Na2SO4, filtered and concentrated. Purification via column chromatography on silica gel (5% ethyl acetate-hexane) afforded olefin 17 (390 mg, 55%).  $[\alpha]_D = -87.3$  (*c* 12.9, CHCl<sub>3</sub>). IR (thin film) 3500 (br), 2930, 1713 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.13 (2H, m), 7.50 (1H, m), 7.40 (2H, m), 5.82 (1H, ddd, J = 17.0, 10.7, 7.8 Hz), 5.14 (1H, m), 5.06 (1H, s), 5.03 (1H, d, J = 8.1 Hz), 4.73 (1H, d, J = 6.3 Hz), 4.65 (1H, d, J = 6.4 Hz), 4.45 (1H, m), 4.18 (1H, dd, *J* = 9.5, 5.0 Hz), 3.88 (1H, m), 3.79 (1H, m), 3.65–3.75 (3H, m), 3.64 (1H, m), 3.58 (3H, s), 3.57 (1H, m), 3.51 (2H, m), 3.37 (3H, s), 2.82 (1H, s br), 2.18 (1H, m), 1.83 (1H, dd, J = 15.0, 4.0 Hz), 1.72-1.79 (3H, m), 1.62 (1H, m), 1.45-1.50 (2H, m), 1.07 (3H, s), 1.02 (3H, d, J = 6.8 Hz), 0.96 (3H, d, J = 7.1 Hz), 0.89 (3H, s), 0.84 (9H, s), 0.01 (3H, s), 0.00 (3H, s).  $^{13}\mathrm{C}$  NMR (125 MHz, CDCl\_3)  $\delta$ 166.1, 140.8, 132.7, 130.7, 129.9, 128.1, 115.1, 106.5, 98.1, 87.9, 83.7, 79.3, 73.1, 71.8, 71.4, 68.0, 64.0, 61.3, 60.1, 58.9, 50.6, 44.4, 36.1, 35.0, 34.8, 27.8, 25.9, 22.8, 18.2, 17.6, 15.9, 10.1, -5.3, -5.4. HRFABMS calcd for (M + Na)  $C_{38}H_{64}O_{10}SiNa$ : 731.4166, found: 731.4205.

(2R,2(15,35,45),3R,5R,75,8R,9R)-9-Benzoyloxy-7-(2-tert-butyldimethylsiloxyethyl)-2-(3-benzoyloxy-1-methoxy-4-methyl-5-hexenyl)-3-(2-methoxyethoxymethyl)oxy-4,4,8-trimethyl-1,6-dioxaspiro[4.5]decane (18). To a solution of alcohol 17 (712 mg, 1.00 mmol, 1 equiv) in pyridine (10 mL) was added DMAP (32 mg, 0.26 mmol, 0.26 equiv), followed by benzoyl chloride (190  $\mu$ L, 1.64 mmol, 1.6 equiv). The clear yellow-brown reaction mixture was heated at 58 °C for 14.5 h, at which time concentration afforded a brown slurry. Purification via column chromatography on silica gel (10-20% ethyl acetate-hexane) afforded dibenzoate 18 (578 mg, 71%), as well as a tribenzoate byproduct (101 mg, 13%).  $[\alpha]_D = -76.0$  (c 12.0, CHCl<sub>3</sub>). IR (thin film) 2930, 1719 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.09 (2H, m), 8.02 (2H, m), 7.51 (1H, m), 7.47 (1H, m), 7.39 (2H, m), 7.35 (2H, m), 5.82 (1H, ddd, J = 17.4, 9.7, 7.1 Hz), 5.48 (1H, m), 5.11 (1H, m), 5.04 (2H, m), 4.71 (1H, d, J = 6.9 Hz), 4.67 (1H, d, J = 6.9 Hz), 4.41 (1H, m), 4.06 (1H, dd, J = 9.1, 4.9 Hz), 3.72 (1H, ddd, J = 9.0, 5.3, 3.4 Hz), 3.63 (1H, ddd, J = 10.1, 6.2, 4.2 Hz), 3.60 (1H, d, J = 4.9 Hz), 3.53 (3H, s), 3.40-3.50 (4H, m), 3.35 (3H, s), 2.53 (1H, m), 1.81 (2H, m), 1.75 (2H, m), 1.60 (1H, m), 1.52 (1H, ddd, J = 12.5, 10.3, 10.3)2.2 Hz), 1.41 (1H, m), 1.07 (3H, s), 1.04 (3H, d, J = 6.9 Hz), 0.92 (3H, d, J = 7.2 Hz), 0.89 (3H, s), 0.71 (9H, s), -0.18 (3H, s), -0.19 (3H, s). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 166.1, 166.0, 139.2, 132.6, 130.7, 130.6, 129.8, 129.5, 128.2, 128.0, 115.7, 106.5, 97.9, 87.6, 84.5, 77.8, 73.8, 73.1, 71.6, 67.9, 64.2, 60.6, 60.5, 58.9, 50.4, 42.5, 35.7, 34.4, 33.8, 27.8, 25.7, 23.0, 17.9, 17.7, 15.7, 10.0, -5.5, -5.6.

(2R,2(1S,3S,4S,5R,6R),3R,5R,7S,8R,9R)-9-benzoyloxy-7-(2-tertbutyldimethylsiloxyethyl)-2-(5-hydroxy-1-methoxy-4,6-dimethyl-3benzoyloxy-8-nonenyl)-3-(2-methoxyethoxymethyl)oxy-4,4,8-trimethyl-1,6-dioxaspiro [4.5]decane (19a). Ozone was bubbled through a solution of olefin 18 (841 mg, 1.03 mmol, 1 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (15 mL) at -78 °C until a light blue color persisted. The excess ozone was purged via a stream of argon (until colorless), at which time DMS (3.6 mL, ~50 equiv) was added. The reaction mixture was warmed to room temperature slowly. After 11 h, triphenylphosphine (105 mg total, ~0.4 equiv) was added in two portions over 3 h. The reaction mixture was concentrated after 16 h (total reaction time), and the resulting crude aldehyde was carried on without purification.

To a cloudy solution of KOtBu (600 mg, 5.35 mmol, 5.2 equiv) and trans-2-butene (~2 mL, excess) in THF (6 mL) at -78 °C was added dropwise 2.02 M nBuLi (2.30 mL, 4.65 mmol, 4.5 equiv). Upon initial addition of *n*BuLi, the reaction mixture became yellow. After, 15 min, a solution of (+)-B-Methoxydiisopinocampheylborane (1.74 g, 5.49 mmol, 5.3 equiv) in THF (2 mL) was added dropwise. Addition of the borane solution resulted in a clear, colorless reaction mixture. After 30 min, BF<sub>3</sub>-Et<sub>2</sub>O (600 µL, 4.86 mmol, 4.7 equiv) was added rapidly, followed immediately by a solution of the crude aldehyde in THF (2.2 mL + 1.2 mL rinse). The resulting cloudy, colorless reaction mixture was stirred at -78 °C for 3.5 h. Then 3 N NaOH (4 mL) and 30% H<sub>2</sub>O<sub>2</sub> (2 mL) were added, and the cold bath was removed. After 1 h, the mixture was diluted with ethyl acetate (20 mL) and poured into brine (20 mL), and the layers were separated. The aqueous layer was extracted with ethyl acetate (5  $\times$  30 mL), and the combined organics were dried over Na2SO4, filtered, and concentrated. The resultant crude was eluted in THF (10 mL), 3 N NaOH (3 mL), and 30% H<sub>2</sub>O<sub>2</sub> (1.5 mL) to complete oxidation of the borane intermediate. After 1 h, the mixture was worked-up as previously mentioned. The excess isopinecamphol was removed via Kugelrohr distillation (high vacuum pressure at 65 °C). Purification via column chromatography on silica gel (5-20% ethyl acetate-hexane) afforded olefins 19a (315 mg, 35%) and 19b (351 mg, 39%) (74% overall for both steps). For **19a**:  $[\alpha]_D = 71.9$  (*c* 10.5, CHCl<sub>3</sub>). IR (thin film) 3500 (br), 2930, 1717, 1453 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.03-8.13 (4H, m), 7.47 - 7.55 (2H, m), 7.35 - 7.42 (4H, m), 5.83 (1H, ddd, J = 17.0, 10.5,8.5 Hz), 5.68 (1H, m), 5.12 (1H, m), 5.06 (2H, m), 4.71 (1H, d, J = 6.7 Hz), 4.68 (1H, d, J = 6.7 Hz), 4.43 (1H, ddd, J = 6.3, 4.1, 2.1 Hz), 4.10 (1H, dd, J = 9.1, 4.9 Hz), 3.69 (2H, m), 3.64 (1H, d, J = 4.8 Hz), 3.60 (3H, s), 3.36-3.58 (6H, m), 3.33 (3H, s), 3.30 (1H, m), 2.60 (1H, br), 2.41 (1H, m), 2.11 (1H, m), 1.57–1.90 (6H, m), 1.45 (1H, m), 1.14 (3H, d, J = 6.9 Hz), 1.09 (3H, s), 0.96 (3H, d, J = 7.3 Hz), 0.94 (3H, d, J = 7.4 Hz), 0.91 (3H, s), 0.75 (9H, s), -0.14 (6H, s). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  166.2, 165.8, 139.2, 132.7, 132.7, 130.8, 130.6, 129.8, 129.5, 128.2, 128.1, 116.0, 106.6, 97.9, 87.5, 84.6, 79.0, 76.6, 73.2, 72.6, 71.7, 68.0, 64.3, 60.8, 60.7, 58.9, 40.6, 39.6, 35.9, 34.6, 31.8, 27.8, 25.7, 23.0, 18.0, 17.8, 17.7, 11.2, 10.0, -5.4, -5.5. HRFABMS calcd for (M + Na) C<sub>48</sub>H<sub>74</sub>O<sub>12</sub>SiNa: 893.4847, found: 893.4817.

(2R,2(15,35,45,55,65),3R,5R,75,8R,9R)-9-benzoyloxy-7-(2-tert-butyl dimethyl siloxy ethyl) - 2 - (5 - hydroxy - 1 - methoxy - 4, 6 - dimethyl - 3 - ben - benzoyloxy-8-nonenyl)-3-(2-methoxyethoxymethyl)oxy-4,4,8-trimethyl-**1,6-dioxaspiro** [4.5]decane (19b).  $[\alpha]_D = -70.3$  (*c* 10.9, CHCl<sub>3</sub>). IR (thin film) 3500 (br), 2930, 1713, 1453 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  8.07 (4H, m), 7.32–7.56 (6H, m), 5.83 (1h, ddd, J = 17.8, 10.3, 7.9 Hz), 5.46 (1H, m), 5.04-5.14 (3H, m), 4.75 (2H, m), 4.40 (1H, ddd, J = 7.6, 5.1, 1.8 Hz), 4.10 (1h, dd, J = 9.0, 4.9 Hz), 3.0.39-3.77 (12H, m), 3.35 (3H, s), 3.34 (1H, m), 2.50 (1H, br), 2.31 (1H, m), 1.97 (2H, m), 1.83 91H, dd, J = 15.2, 4.2 Hz), 1.76 (3H, m), 1.57 (1H, m), 1.41 (1H, m), 1.09 (3H, s), 0.97 (3H, d, J = 6.9 Hz), 0.93 (3H, d, J = 6.8 Hz), 0.93 (3H, d, J = 7.1 Hz), 0.91 (3H, s), 0.71 (9H, s), -0.17 (3H, s), -0.19 (3H, s). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 166.7, 166.1, 141.6, 132.9, 132.6, 130.6, 130.3, 129.8, 129.6, 128.3, 128.0, 115.3, 106.6, 97.8, 87.3, 84.5, 78.0, 74.1, 73.8. 71.7, 68.1, 64.2, 60.6, 60.5, 59.0, 50.4, 40.9, 39.4, 35.6, 34.3, 33.6, 27.8, 25.7, 23.0, 17.9, 17.7, 16.4, 10.0, 8.6, -5.5, -5.6. HRFABMS calcd for (M + Na) C48H74O12SiNa: 893.4847, found: 893.4860.

(2R,2(1S,3S,4S,5R,6R),3R,5R,7S,8R,9R)-9-tert-Butyldimethylsiloxy-7-(2-tert-butyldimethylsiloxyethyl)-2-(1-methoxy-4,6-dimethyl-3,5bis(tert-butyldimethylsiloxy)-8-nonenyl)-3-(2-methoxyethoxymethyl)oxy-4,4,8-trimethyl-1,6-dioxaspiro[4.5]decane (21).  $[\alpha]_D = -41.9$ (c 1.7, CHCl<sub>3</sub>). IR (thin film) 2928, 1472 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz,  $CDCl_3$ )  $\delta$  5.89 (1H, ddd, J = 17.4, 10.4, 7.1 Hz), 4.98 (2H, m), 4.80 (1H, d, *J* = 7.3 Hz), 4.57 (1H, d, *J* = 7.3 Hz), 4.38 (1H, m), 4.30 (1H, m), 3.96 (1H, dd, J = 9.1, 4.7 Hz), 3.75 - 3.85 (3H, m), 3.62 (3H, s), 3.47-3.67 (6H, m), 3.38-3.41 (1H, m), 3.38 (3H, m), 2.42 (1H, m), 1.86 (1H, m), 1.71 (1H, m), 1.64 (1H, dd, J = 14.2, 4.0 Hz), 1.40-1.53 (3H, m), 1.25 (2H, m), 1.05 (3H, s), 1.04 (3H, d, J = 6.9 Hz), 0.91 (3H, s), 0.89 (9H, s), 0.88 (9H, s), 0.87 (18H, s), 0.81 (3H, d, J = 7.1 Hz), 0.74 (3H, d, J = 7.1 Hz), 0.13 (3H, s), 0.10 (3H, s), 0.07 (3H, s), 0.05 (3H, s), 0.04 (3H, s), 0.03 (6H, s), 0.01 (3H, s). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 141.5, 113.6, 106.8, 98.7, 89.6, 86.4, 77.6, 77.4, 71.8, 71.4, 68.1, 67.8, 64.0, 61.3, 60.3, 59.0, 50.1, 43.7, 42.6, 37.9, 36.1, 33.9, 31.2, 29.7, 26.3, 26.1, 26.0, 25.9, 25.7, 23.3, 18.4, 18.3, 18.2, 18.1, 17.9, 15.4, 10.6, 10.4, -3.0, -3.8, -4.0, -4.0, -4.6,-5.0 (3 carbons).

(2R,3S,4S)-N-[(1R,3S)-3-[4-(1E)-3[(2R,3R,5R,7S,8R,9R)-2-[(1S,-1)-2-[(1S3S,4S,5R,6R,7E,9E,11E,13Z)-3,5-Bis(tert-butyldimethylsiloxy)-14-cyano-1-methoxy-4,6,8,9,13-pentamethyl-7,9,11,13-tetradecatetraenyl]-9-(tert-butyldimethylsiloxy)-3-[bis(p-methoxybenzyl)phosphatyl]-4,4,8-trimethyl-1,6-dioxaspiro[4.5]dec-7-yl]propenyl]-2-oxazolyl]-1methylbutyl]-2,3-dihydroxy-4-(dimethylamino)-5-methoxy Valeramide (28). To a solution of aldehyde 26 (9.6 mg, 0.015 mmol) and phosphonium salt 27 (10.4 mg, 0.008 mmol) in DMF (200 µL) at 0 °C was added LDA (32 µL, 0.5 M in THF, 1 equiv). Four successive portions of LDA (3  $\times$  32  $\mu$ L, then 15  $\mu$ L, 0.5 M in THF) were added over 30 min, at which time the reaction mixture was partitioned between ethyl acetate (10 mL) and saturated aqueous NaHCO<sub>3</sub> (10 mL). The layers were separated, and the aqueous layer was extracted with ethyl acetate (3  $\times$  10 mL). The combined organics were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. Purification via column chromatography on silica gel (2-6% CH<sub>3</sub>OH-CH<sub>2</sub>Cl<sub>2</sub>) afforded aldehyde 26 (6 mg, 58%) and olefin 28 (3 mg, 23%, 55% based on recovered starting material, **26**). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.36 (1H, s), 6.99–7.08 (4H, m), 6.77–6.85 (5H, m), 6.36 (1H, d, *J* = 11.1 Hz), 6.16 (2H, m), 6.07 (1H, d, J = 8.9 Hz), 5.06 (1H, s), 5.01 (1H, dd, J = 20.9, 11.6 Hz), 4.91 (1H, dd, J = 25.6, 13.6 Hz), 4.80 (1H, dd, J = 11.2, 5.3 Hz), 4.67 (1H, dd, J = 11.0, 5.8 Hz), 4.56 (2H, m), 4.38 (1H, br d), 4.29 (1H, br), 4.09 (3H, m), 3.84 (1H, br), 3.76-3.79 (9H, m), 3.66 (4H, m), 3.58 (1H, m), 3.48 (1H, m), 3.35 (3H, s), 2.99 (1H, m), 2.73 (7H, br m), 2.65 (2H, m), 2.07 (3H, s), 2.01 (3H, s), 1.88 (3H, s),

1.40-1.85 (8H, m), 1.33 (3H, d, J = 6.8 Hz), 1.25 (3H, s), 1.19 (3H, d, J = 6.5 Hz), 1.16 (3H, s), 1.03 (3H, d, J = 6.9 Hz), 0.94 (9H, s), 0.88 (3H, d, J = 9.0 Hz), 0.86 (9H, s), 0.85 (9H, s), 0.82 (3H, d, J = 7.0 Hz), 0.20 (3H, s), 0.08 (3H, s), 0.07 (3H, s), 0.05 (3H, s), 0.04 (3H, s), -0.01 (3H, s).

34R-Calyculin C (29). To a Nalgene tube (2 mL) containing protected calyculin 28 (3 mg, 1.8  $\mu$ mol) and a magnetic stirbar was added an HF stock solution (130  $\mu$ L; composed of 215  $\mu$ L of CH<sub>3</sub>CN, 45  $\mu$ L of H<sub>2</sub>O, and 23  $\mu$ L of 48% aqueous HF). After a total reaction time of 65 h, the entire reaction mixture was chromatographed directly on silica gel (0-12% CH<sub>3</sub>OH-CH<sub>2</sub>Cl<sub>2</sub>) to afford 34R-calyculin C 29 (1 mg, 52%). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.34 (1H, m), 7.33 (1H, s), 6.99 (1H, dd, J = 14.9, 11.1 Hz), 6.88 (1H, m), 6.81 (1H, d, J = 14.9 Hz), 6.32 (1H, d, J = 11.5 Hz), 6.13 (1H, d, J = 16.1 Hz), 6.01 (1H, J = 9.8 Hz), 5.05 (1H, s), 4.57 (1H, m), 4.45 (1H, s), 4.35 (1H, s)m), 4.26 (1H, dd, J = 10.3, 3.5 Hz), 4.24 (1H, m), 4.05 (1H, m), 3.97 (2H, m), 3.87 (1H, m), 3.78 (1H, dd, J = 9.7, 9.4 Hz), 3.56 (2H, m), 3.54 (3H, s), 3.39 (3H, s), 3.16 (1H, m), 2.85 (6H, s), 2.73 (1H, m), 2.49 (1H, m), 2.07 (3H, d, J = 1.0 Hz), 1.97 (3H, s), 1.84 (3H, s), 1.45-2.10 (8H, m), 1.32 (3H, d, J = 6.6 Hz), 1.29 (3H, d, J = 6.9Hz), 1.23 (3H, s), 1.03 (3H, d, J = 6.9 Hz), 0.95 (3H, s), 0.88 (3H, d, J = 7.1 Hz), 0.71 (3H, d, J = 6.8 Hz). HRFABMS calcd for C<sub>51</sub>H<sub>84</sub>N<sub>4</sub>O<sub>15</sub>P (MH<sup>+</sup>): 1023.5671, found: 1023.5690.

(2S,3S,4S)-N-[(1R,3S)-3-[4-(1E)-3](2R,3R,5R,7S,8R,9R)-2-[(1S,3S,-4S,5R,6R,7E,9E,11E,13Z)-3,5-Bis(tert-butyldimethylsiloxy)-14-cyano-1-methoxy-4,6,8,9,13-pentamethyl-7,9,11,13-tetradecatetraenyl]-9-(tert-butyldimethylsiloxy)-3-[bis(p-methoxybenzyl)phosphatyl]-4,4,8trimethyl-1,6-dioxaspiro[4.5]dec-7-yl]propenyl]-2-oxazolyl]-1methylbutyl]-2,3-dihydroxy-4-(dimethylamino)-5-methoxy Valeramide (30). To a solution of aldehyde 26 (9 mg, 0.007 mmol) and phosphonium salt 3 (10 mg, 0.017 mmol) in DMF (120 µL) at 0 °C was added LDA (37 µL, 0.5M in THF, 1 equiv). Six successive portions of LDA (6  $\times$  37  $\mu$ L, 0.5M in THF) were added over 1 h, at which time the reaction mixture was partitioned between ethyl acetate (10 mL) and saturated aqueous NaHCO3 (10 mL). The layers were separated, and the aqueous layer was extracted with ethyl acetate (5  $\times$ 10 mL). The combined organics were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. Purification via column chromatography on silica gel (2-6% CH<sub>3</sub>OH-CH<sub>2</sub>Cl<sub>2</sub>) afforded aldehyde 26 (4 mg, 44%) and olefin 30 (2.7 mg, 24%, 41% based on recovered starting material, 26). HRFABMS calcd for C<sub>85</sub>H<sub>142</sub>N<sub>4</sub>O<sub>17</sub>PSi<sub>3</sub> (MH<sup>+</sup>): 1605.9415, found: 1605.9381

**Calyculin C (1).** To a Nalgene tube (2 mL) containing protected calyculin **30** (2.5 mg, 1.5  $\mu$ mol) was added an HF stock solution (100  $\mu$ L; composed of 215  $\mu$ L of CH<sub>3</sub>CN, 45  $\mu$ L of H<sub>2</sub>O, and 23  $\mu$ L of 48% aqueous HF). After a total reaction time of 60 h, the entire reaction mixture was chromatographed directly on silica gel (0–5% CH<sub>3</sub>OH–CH<sub>2</sub>Cl<sub>2</sub>) to afford calyculin C (1) (0.4 mg, 30%). Thin-layer chromatography of 1 was performed in multiple solvent systems against an authentic sample of calyculin C.<sup>17</sup> For <sup>1</sup>HNMR of 1 and authentic calyculin C, see Supporting Information. HRFABMS calcd for C<sub>51</sub>H<sub>84</sub>N<sub>4</sub>O<sub>15</sub>P (MH<sup>+</sup>): 1023.5671, found: 1023.5723.

Acknowledgment. This paper is dedicated to Professor Yoshito Kishi on the occasion of his 60th birthday. This work was supported by funding from the NIH (Grant No. 51095). The authors are deeply indebted to past group members, without whose efforts this work would not be possible. The authors thank Varsha Gupta for her efforts in providing the  $C_{33}$ - $C_{37}$  fragment. The authors graciously thank Professor Nobuhiro Fusetani for his generous gift of authentic calyculin C.

**Supporting Information Available:** Experimental procedures and/or data for compounds **8**, **10–12**, and **22**, as well as copies of <sup>1</sup>H NMR for compounds **1**, **5–19b**, **21–22**, and **28–29** are provided. Mass spectral data for compounds **1** and **30** are also provided (36 pages, print/PDF). See any current masthead page for ordering information and Web access instructions.

JA980836Q